

# **XpressRNA Viral Kit**

Protocol for isolation of Total Viral RNA from oral and nasopharyngeal swabs collected in Viral Transport Medium (VTM) or Molecular Transport Medium (MTM).

# **Process Flow**



# **Kit Contents**

Components	Storage Conditions	Shipping Conditions
Viral Lysis Buffer	RT	RT
Viral MagNa Mix	2 - 8 °C	RT
Viral Wash Buffer	RT	RT
Viral Elution Buffer	RT	RT
MagNa Stand (optional)	RT	RT
Carrier RNA	-20°C	RT
Carrier RNA dilution buffer	-20°C	RT

\*RT denotes 15 - 25°C.

#### Materials not provided with the kit

- 1. 100% Ethanol to Wash Buffers as indicated on the bottle.
- 2. Water bath/heat block at 56°C.
- Reconstitute Carrier RNA with Carrier RNA dilution buffer and store at -20°C (Stock concentration: 1 mg/ml).

#### Important

Pay attention to standard lab practices and safety information before beginning the procedure. For more information, refer the appropriate Material Safety Data Sheet (MSDS) available from the product supplier or download from our website <u>http://www.maggenome.com/</u>

### **Technical Support**

For any product related queries please write to us on <u>info@maggenome.com</u>, <u>sales@maggenome.com</u>, <u>support@maggenome.com</u>.



Protocol		
Lysate Preparation	1. Take 200 μl of VTM/fluid sample in a 1.5 ml tube.	
	Add 100 $\mu$ l of Viral Lysis Buffer and pipette mix the contents thoroughly.	
	3. Add <b>5 µl of Carrier RNA</b> and pipette mix.	
	4. Vortex the tube for 30 seconds.	
	5. Briefly spin the tube to bring down the residues from the inside of the lid.	
	6. Incubate for 10 minutes at 56°C.	

	7.	<ul> <li>(Note: Vortex the Viral MagNa Mix to ensure complete dispersion of the particles.)</li> <li>7. Add 125 μl of Viral MagNa Mix to the lysate and gently invert the tube 10 - 15 times</li> </ul>	
		mix properly. Do not vortex.	
	8.	Incubate at RT for 5 minutes.	
<b>RNA</b> Binding	9.	Place the tube on Magna Stand until the solution becomes clear.	
	10.	Carefully discard the supernatant completely without removing the tube from the	
		MagNa Stand. Ensure the magnetic nanoparticles are not disturbed.	

	11.	11. Add 250 µl of Viral Wash Buffer and gently invert the tube for 10 - 15 times with	
		removing the tube from MagNa Stand (surface wash only).	
	12.	Discard the supernatant without removing the tube from MagNa Stand.	
RNA Washing	13.	Repeat the steps 11 - 12.	
	14.	Air dry the magnetic nanoparticles without removing the tube from the MagNa Stand	
		at RT for 2 minutes without over drying them.	

	15.	After drying, remove the tube from the MagNa Stand.
	16.	Add $30 \ \mu l$ of Viral Elution buffer and completely resuspend the magnetic nanoparticles
RNA Elution		by pipette mixing (10 – 15 times).
	17.	Incubate at 56°C for 2 minutes with intermittent tapping.
		(Note: Briefly spin the tube for 5 seconds before placing on the MagNa stand.)
	18.	Place the tube on MagNa Stand for 5 minutes or until the solution appears clear (place
		the magnetic stand on ice).
	19.	Carefully transfer the supernatant containing RNA to a nuclease free 1.5 ml tube
		without removing the tube from MagNa Stand. Ensure the magnetic nanoparticles
		are not disturbed.
	20.	Discard the magnetic nanoparticles in the appropriate hazard container.



# Troubleshooting Guide

Observation	Possible causes	Suggested Solution
		XpressRNA kit buffers are tested and guaranteed
	RNase contamination	RNase-free, although RNase can be introduced during
Degraded DNA		use. Be certain not to introduce RNase during storage
Degraded RNA		and handling.
	Common DNA stores as	Ensure that carrier RNA and dilution buffer are stored
	Callier KINA Stolage	properly at the recommended storage temperature.
		Make sure that the incubation temperature and time
	Incomplete Lysis	for lysis is followed as per the protocol.
	MagNa Mix was	Ensure proper dispersion of nanoparticles by
	improperly handled	vortexing the MagNa Mix prior to use.
Low RNA yield or	Magnetic nanoparticle loss	Carefully remove the supernatant from the tube
Poor Quality	during binding or washing	without removing the tube from the MagNa Stand
	steps	and without disturbing the magnetic nanoparticles.
	Ethanol is not added to wash	Add 100% ethanol to wash buffer before use as
	buffer	indicated on the bottles.
	Improper elution	Completely resuspend the magnetic nanoparticles in
		elution buffer before incubation at 56°C for elution.
Poor performance		Air dry the magnetic papoparticles after the washing
of extracted RNA	Ethanol carryover	steps to remove ethanol traces completely, but do not
in downstream		over dry the pellet
applications		over dry the penet.